

Introduction

Optical modulation of ion channels is traditionally studied using a manual patch clamp system combined with a light source. This approach, however, is limited by a very low throughput. In the present work we show data recorded using a 384-well based automated patch clamp system equipped with 384 integrated light sources (Qube Opto 384).

Optogenetics

In this study we evaluated Channelrhodopsin 2 (ChR2), a light-sensitive non-selective cation channel permeable to Na⁺, K⁺ and Ca²⁺ opened upon illumination (Berndt et al., 2012). Furthermore, we employed the chloride-conducting channelrhodopsin (iC⁺⁺, Govorunova et al., 2015), which was developed from a non-selective cation conducting channelrhodopsin through a mutational approach.

Optopharmacology

Compound activation by light enables the pharmacological manipulation of receptors, ion channels and other proteins with a high degree of temporal control. We used caged GABA (Rubi-GABA, Zayat et al., 2003) to study the light activation of ligands, in combination with the microfluidic flow channel of the QChip 384, to give a higher degree of experimental control.

Concept System

- Full scale, integrated HTS system
 - Optical stimulation combined with APC – fully integrated
 - 384 format
 - Optogenetics module fully controlled by Qube software
 - Dark cabinet for automated handling of light sensitive compounds and cells
 - Compatibility with other Qube 384 features such as temperature control

- Overall features of the optical system
 - Addressing 384 sites
 - Individual control of 24 columns
 - Software controlled
 - ◊ Timing, duration and intensity
 - ◊ Ramps and other optical waveforms

Methods

ChR2/HEK 293, GABA_A($\beta_2\beta_3\gamma_2$)/HEK 293, and iC⁺⁺/HEK 293 cells were cultured according to the supplier's description. ChR2 and GABA_AR were stably expressed in the HEK cells whereas iC⁺⁺ was induced 24 h prior to the experiment. All experiments were carried out at ambient temperature using QChip 384X multi-hole consumables and patched using a standard whole cell protocol.

Extracellular solution:

CaCl₂ 2 mM, MgCl₂ 1 mM, HEPES 10 mM, KCl 4 mM, NaCl 145 mM, Glucose 10 mM, pH = 7.4 with NaOH, osmolality = 305 mOsm with sucrose (before adjustment 285 – 295 mOsm)

Intracellular solution for ChR2 experiments:

CsF 140 mM, EGTA/CsOH 1 mM / 5 mM, HEPES 10 mM, NaCl 10 mM, pH = 7.3 with 3 M CsOH, osmolality = 320 mOsm with sucrose

Intracellular solution for Rubi-GABA and iC⁺⁺ experiments:

KCl 90 mM, KF 50 mM, MgCl₂ 1 mM, HEPES 10 mM, EGTA 11 mM, Mg-ATP 4 mM, pH = 7.35 with KOH, osmolality = 300 mOsm (before adjustment 280 – 290 mOsm)

Conclusions

With the new optical feature of Qube Opto 384, it was possible to evaluate both light activated ion channels and photoactivated ligands:

Qube Opto 384 is a Qube with 384 individual, built-in light sources (currently at 475 nm) that can modulate ion channels through optogenetic actuators (e.g. channelrhodopsin) or activate compounds (caged compounds).

References

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4. Rial Verde EM, Zayat L, Etchenique R, Yuste R.: " Photorelease of GABA with Visible Light Using an Inorganic Caging Group. *Front Neural Circuits.* 2008 Aug 13;2:2. eCollection 2008.

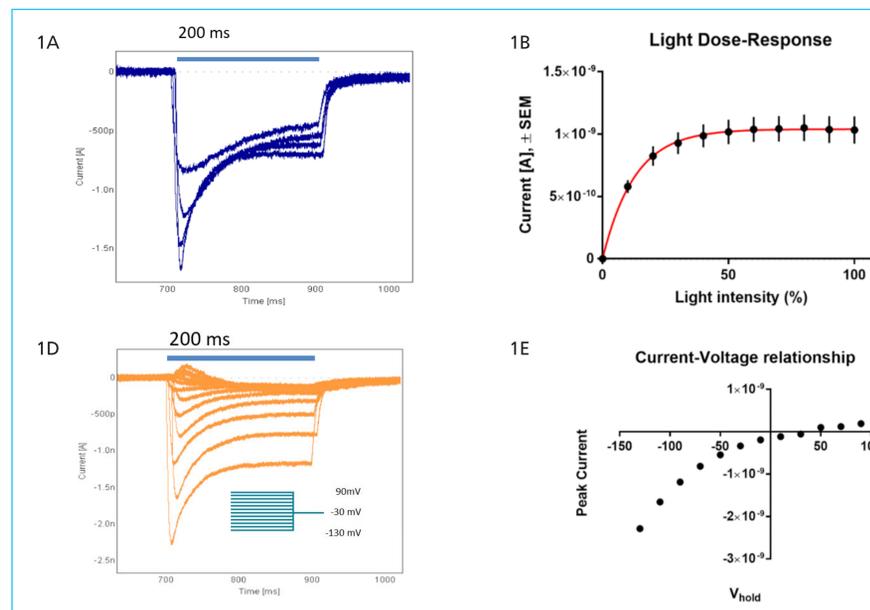


Fig. 1: The ChR2 mediated current could be manipulated in both a light and voltage dependent manner.

A) Typical light-evoked current at a light output intensity of 10%, 20%, 40% and 100% and a holding potential of -90 mV. B) Dose-response relationship for light from 0% to 100% output intensity. C) Fast activation kinetics: Rise time calculated as time from 10 – 90% of maximum current amplitude. Data is represented as mean \pm SD of n=12. C) ChR2: Left: Representative light-evoked current traces B) Current-Voltage relationship. The maximum current amplitude plotted against holding potential.

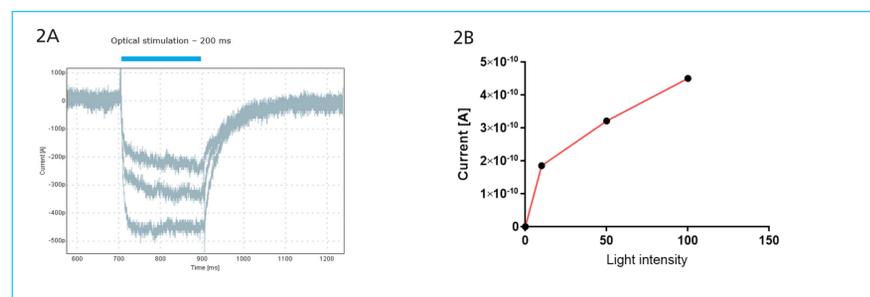


Fig. 2: Chloride conducting channelrhodopsin (iC⁺⁺)

A) iC⁺⁺ sweep plots when exposed to light output intensities of 10%, 50% and 100%. The holding potential was clamped at -100 mV. B) Dose-response relationship for the peak current of the traces seen left.

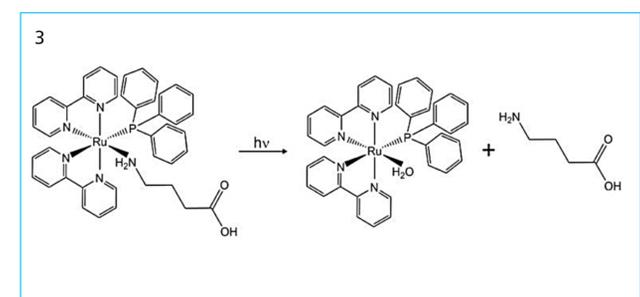


Fig. 3: Rubi-GABA is a caged GABA compound activated by visual wavelengths (Rial Verde et al., 2008).

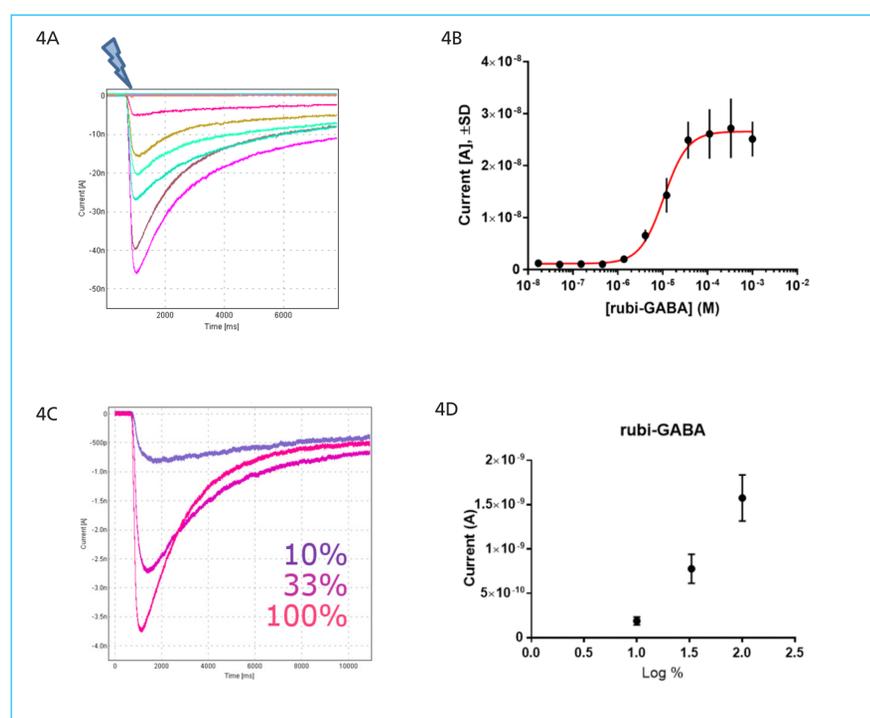


Fig. 4: Photorelease of caged GABA

A) Concentration-response relationship: HEK 293 cells expressing GABA_A($\alpha_5\beta_3\gamma_2$) were exposed to varying concentrations of Rubi-GABA (Tocris), by a 200 ms light pulse (475 nm) at 100% light output. B) Non-cumulative concentration-response curve for Rubi-GABA of a 3-fold dilution series. C) Optical dose-response relationship: GABA_A($\alpha_5\beta_3\gamma_2$) expressing cells were exposed to 1 mM Rubi-GABA and GABA was photoreleased. Light pulse: 200 ms D) Intensity-response relationship for Rubi-GABA at intensities of 10%, 33% and 100% light output. Data is represented as mean \pm SD of n=32.

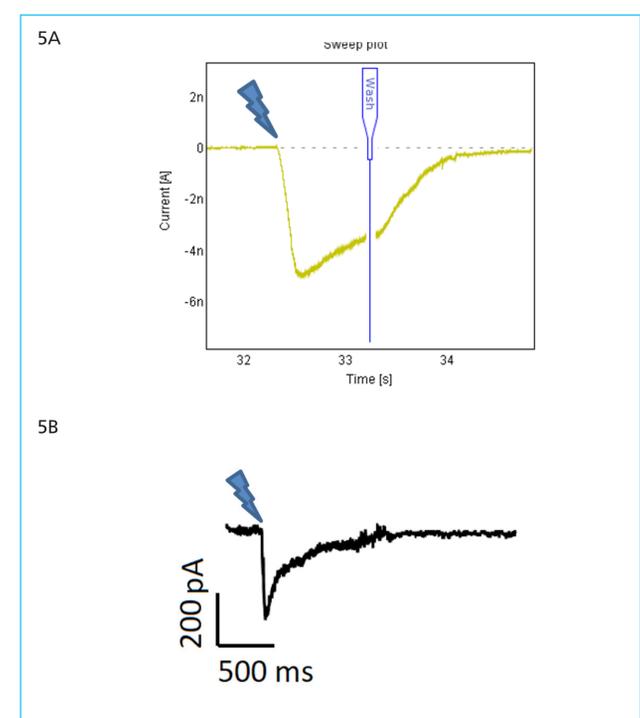


Fig. 5: Fast ligand exposure

With the new optical capability of Qube Opto 384 in combination with the microfluidic system of the QChip 384X, the ligand exposure time can be better controlled and drastically reduced. A) Rubi-GABA photoactivation followed by wash-out (200 μ M, 100 ms light pulse). B) Uncaging during perfusion: 330 μ M Rubi-GABA was washed in and during perfusion, GABA was uncaged by a 20 ms light pulse, resulting in an instantaneous washout of uncaged GABA.