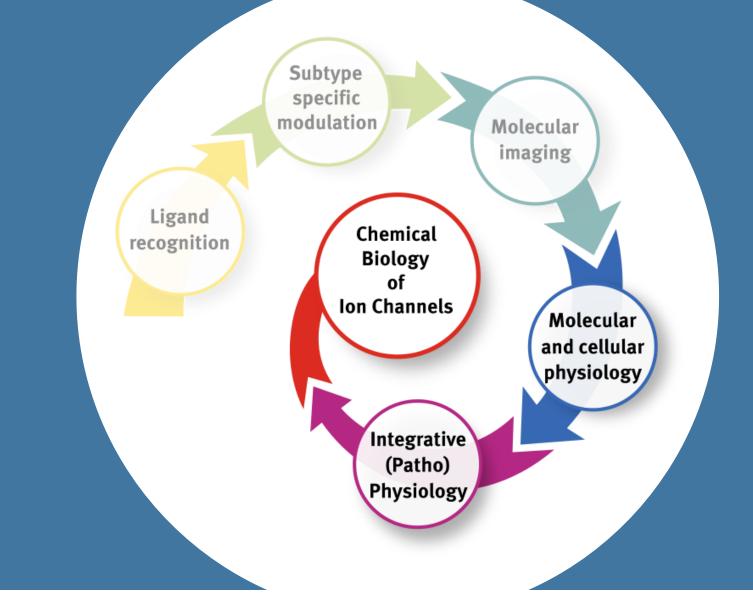


# Analyzing the developmental profile and physiological function of neuronal K<sub>v</sub>7 channels

Elif Karabatak, Thomas Budde
University Hospital Münster, Germany
Institute of Physiology I



#### Introduction

 $K_v7$  (KCNQ or M-) channels generate voltage dependent K<sup>+</sup> currents and represent the molecular basis of the M-current ( $I_M$ ). The five known subtypes ( $K_v7.1-7.5$ ) are characterized by different tissue expression and physiological properties.<sup>[1]</sup> In most neurons, native M-channels are composed of  $K_v7.2$  and  $K_v7.3$  subunits, and are responsible for controlling the neuronal excitability.<sup>[2]</sup> During physiological as well as pathophysiological conditions (e.g. epilepsy) channel modulators play a significant role.<sup>[3]</sup> Previous studies from our group point out the importance of M-channels in the thalamocortical system, making them interesting targets for further investigation.<sup>[4]</sup>

With this project, we thus aim to investigate:

1) The developmental expression profile of  $K_v$ 7.2 and  $K_v$ 7.3 in thalamic neurons. 2) The modulation of these channels by different compounds including  $PIP_{2,}$  cholesterol analogues and novel activators. 3) The properties of  $I_M$  in prethalamic/ thalamic neurons in health and disease.

# Developmental expression profile of $K_v7.2$ and $K_v7.3$ in mouse brain

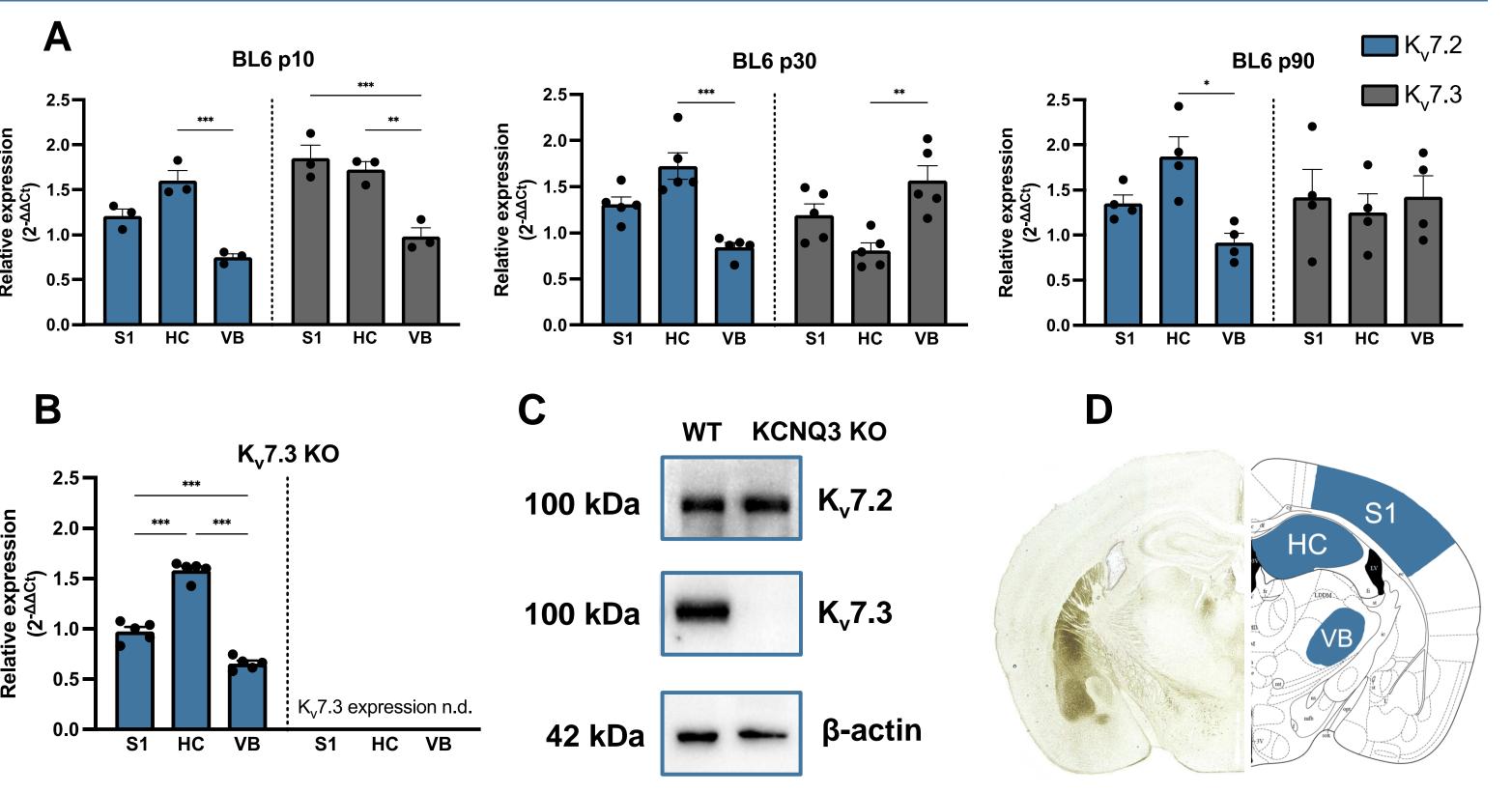


Figure 1:  $K_v$ 7.2 and  $K_v$ 7.3 gene and protein expression in mice during development. S1: Somatosensory cortex. HC: Hippocampus. VB: Ventrobasal complex. **A)** Relative gene expression in C57BL/6JOlaHsd mice. Early infant mice (n=3): p10, adolescent mice (n=4): p30, adult mice (n=4): p90. **B)** Relative gene expression in KCNQ3 KO mice (n=5): p90 and older. One-way Anova test was applied. Error bars represented as  $\pm$  SEM. \*p = 0.033, \*\*p = 0.01, \*\*\*p < 0.001. **C)** Representative data for western blotting.  $K_v$ 7.2 and  $K_v$ 7.3 in mouse brain: p30. **D)** Regions of interest in coronal brain slices

linker

# Error bars represented as $\pm$ SEM. \*p = 0.033, \*\*p = 0.01, \*\*\*p < 0.001. We western blotting. K<sub>v</sub>7.2 and K<sub>v</sub>7.3 in mouse brain: p30. **D)** Regions of inter (Modified from mouse brain atlas, Paxinos and Franklin). Cell transfection

KCNQ2

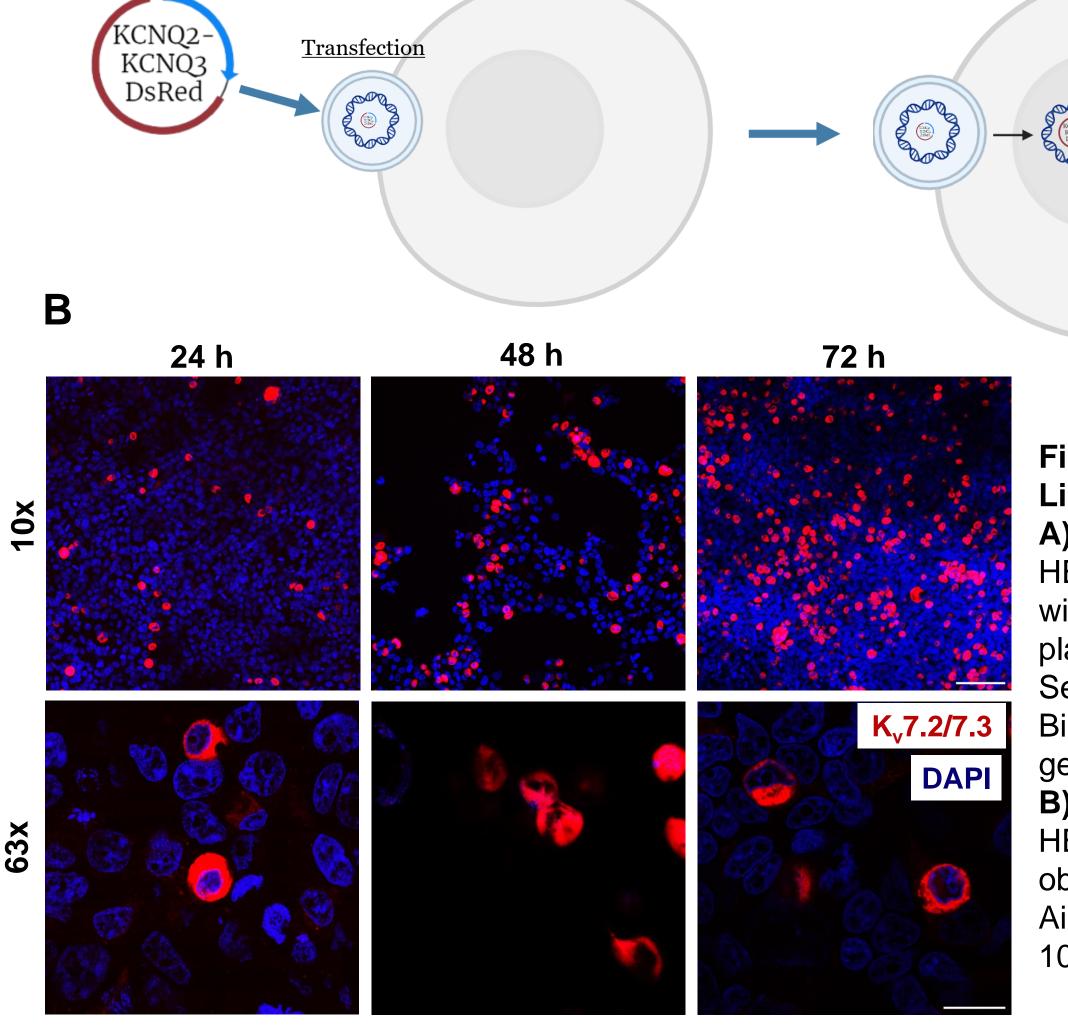


Figure 3: Cell transfection with Lipofectamine™ 3000.

Expression of functional

Kv7.2/7.3

A) Schematic representation of HEK-293FT cell line transfection with the KCNQ2-KCNQ3-DsRed plasmid, designed by Prof. Guiscard Seebohm, and obtained from BioCat GmbH, Heidelberg. Figure generated with BioRender.

**B)** Post-transfection imaging of HEK-293FT cells. Images were obtained with Zeiss LSM 900, Airyscan. Scale bars: 100 μM for 10x, 20 μM for 63x.

#### Methods

for the chart 1) RT-qPCR

for the investigation of ion channel expression on mRNA level

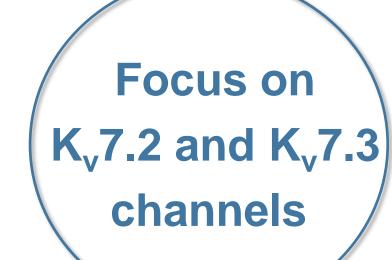
#### 5) Patch-clamp

for the examination of electrophysiological ion channel activity



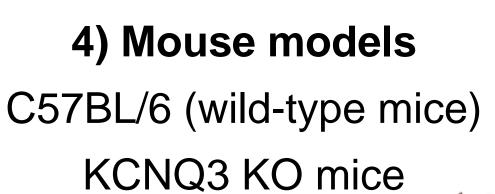
#### 2) Western blot

for the investigation of ion channel expression on protein level



3) Cell transfection

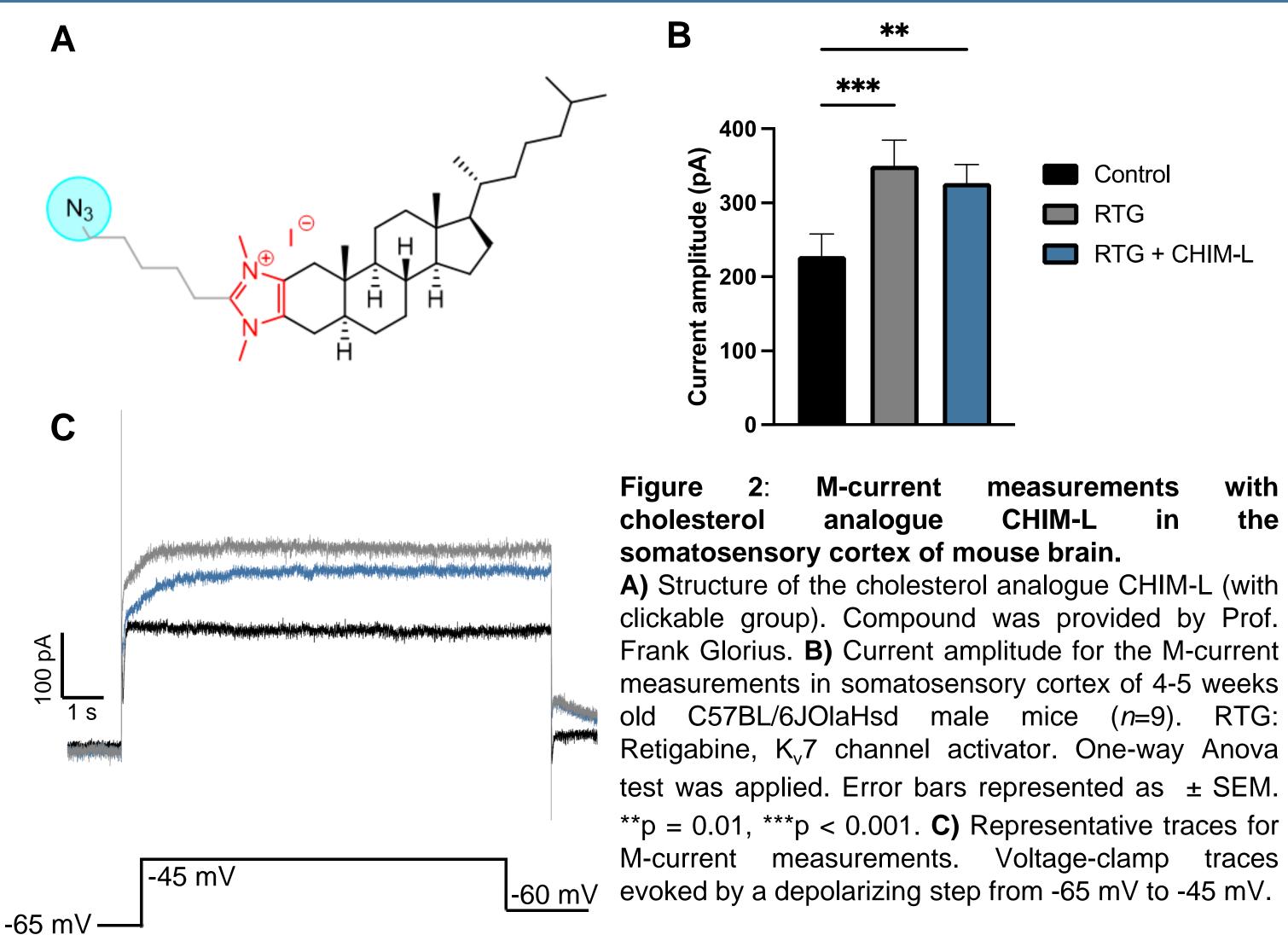
for generating a cell line with functional M-channel





Figures generated with BioRender

# $K_{\nu}7$ modulation with the cholesterol analogue CHIM-L



## Functionality test of transfected cells

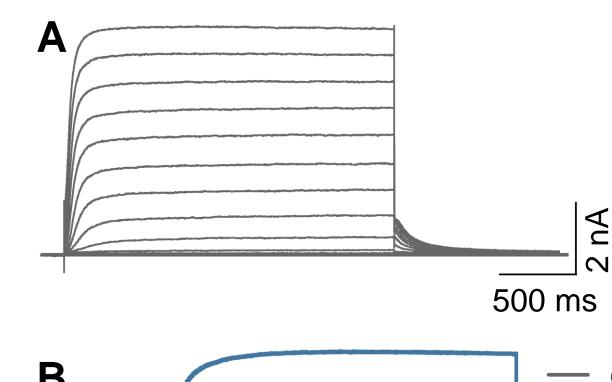
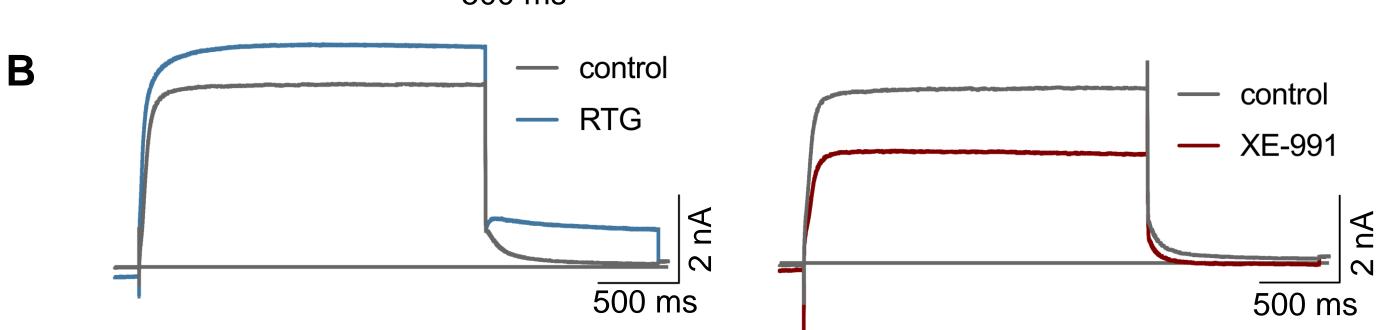


Figure 4: M-Channel modulation
Representative traces of transfected cell measurements.
Currents were evoked by 2 s depolarising steps to test potentials ranging from -80 to +40 mV in 10 mV increments from a holding potential of -80 mV in 15 s intervals. Tail currents were recorded at -60 mV. A) Control measurement. B) Control measurement in comparison to M-channel activator retigabine (RTG, 10 μM) and M-channel blocker XE-991 (10 μM).



## Future prospects

Test of M-channel modulators with patch-clamp

- Natural cholesterol
- Cholesterol analogues (CHIM-L)
- PIP<sub>2</sub> analogues
- Retigabine analogues

