

Validation of K_v7.X channel assays using the Qube 384 automated patch-clamp platform



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1 ABSTRACT

K_v7 (KCNQ) channels are voltage-gated K⁺ channels with major roles in neurons, muscle cells and epithelia. The biophysical properties of K_v7 channels make them essential in controlling the activity of excitable cells¹. They produce currents that are voltage-activated, slowly activating and non-inactivating, such as neuronal M current and cardiac I_{Ks}². These channels often work as 'excitability brakes'³ and are targeted by various hormones and modulators to regulate cellular activity outputs. Genetic deficiencies in KCNQ genes result in human excitability disorders, including epilepsy, arrhythmias and deafness⁴, making K_v7 channels an attractive target for the pharmaceutical research. The aim of the present work was to demonstrate using the Sophion Qube 384-well APC platform for robust and reproducible K_v7 currents recordings, suitable for fast and reliable drug testing.

3 RESULTS

3.1 Biophysical properties of K_v7 cell lines on the Sophion Qube

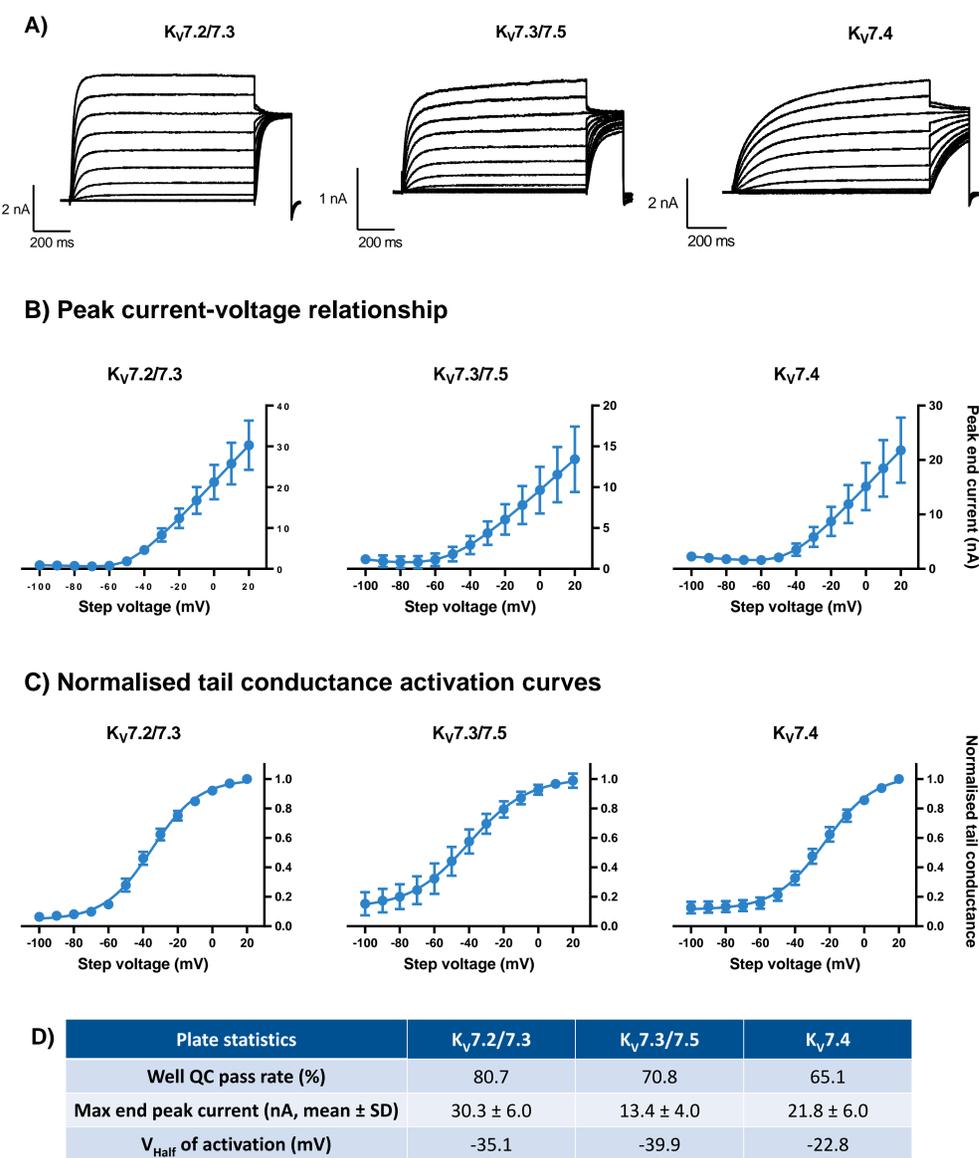


Figure 1. Representative K_v7 currents, IV and normalised tail conductance activation curves, with plate statistics used for assay QC. Representative current traces (A), current-voltage (IV, B) and normalised tail conductance activation curves (C) for K_v7.2/7.3, K_v7.3/7.5 and K_v7.4. Data are plotted as mean ± SD, n ≥ 250. Plate statistics used for assay quality control represented in (D).

4 SUMMARY

K_v7.2/7.3, K_v7.3/7.5 and K_v7.4 can be assessed by automated patch clamp on the Sophion Qube platform. Biophysical characterization shows robust current amplitudes and V_{Half} of activation, which correspond to those reported in the published literature, with good assay performance. Pharmacological tool compounds with different mechanisms of action, both activators and inhibitors, were profiled in concentration response curves. Some compounds show subtype selectivity, for example, QO 58 shows stronger activation of K_v7.4 compared to K_v7.2/7.3 and K_v7.3/7.5, shown in both delta V_{Half} and % activation at the -40 mV step voltage. ML 252 shows strong inhibition of all K_v7 subtypes at the 20 mV step voltage, whilst only alters the V_{Half} of activation in the K_v7.2/7.3 subtype. Overall, these results demonstrate a robust screening methodology to detect compounds which inhibit and activate K_v7.X channels through both alterations in the V_{Half} of activation and current amplitude, using the Sophion Qube platform.

Acknowledgement

Thank you to the cell culture team at Charles River for their help and support.

2 MATERIAL AND METHODS

Cell Culture: K_v7.2/7.3, K_v7.3/7.5 and K_v7.4 cells were produced at Charles River Laboratories and are commercially available.

Qube experiments: Experiments were conducted using Multi-hole QChips on the Sophion Qube 384 platform, using EC000 and modified IC000 as prescribed by Sophion. Cells were held at -80 mV throughout the experiment. K_v7.x currents were recording with 1 s voltage steps from -100 to +20 mV. Currents were sampled at 25 kHz, with a cut off at 5 kHz and Bessel filtering. QC filters used: RSeal > 3 or 4 MΩ, Capacitance > 20 pF, resting peak current > 1.5 or 3 nA.

Analysis: Data analysis was performed using Qube Analyzer and GraphPad Prism (9.0). Compound effects on the V_{Half} of activation and current size at -40 and +20 mV voltage steps were normalised to vehicle application.

3.2 K_v7 pharmacology of reference compounds

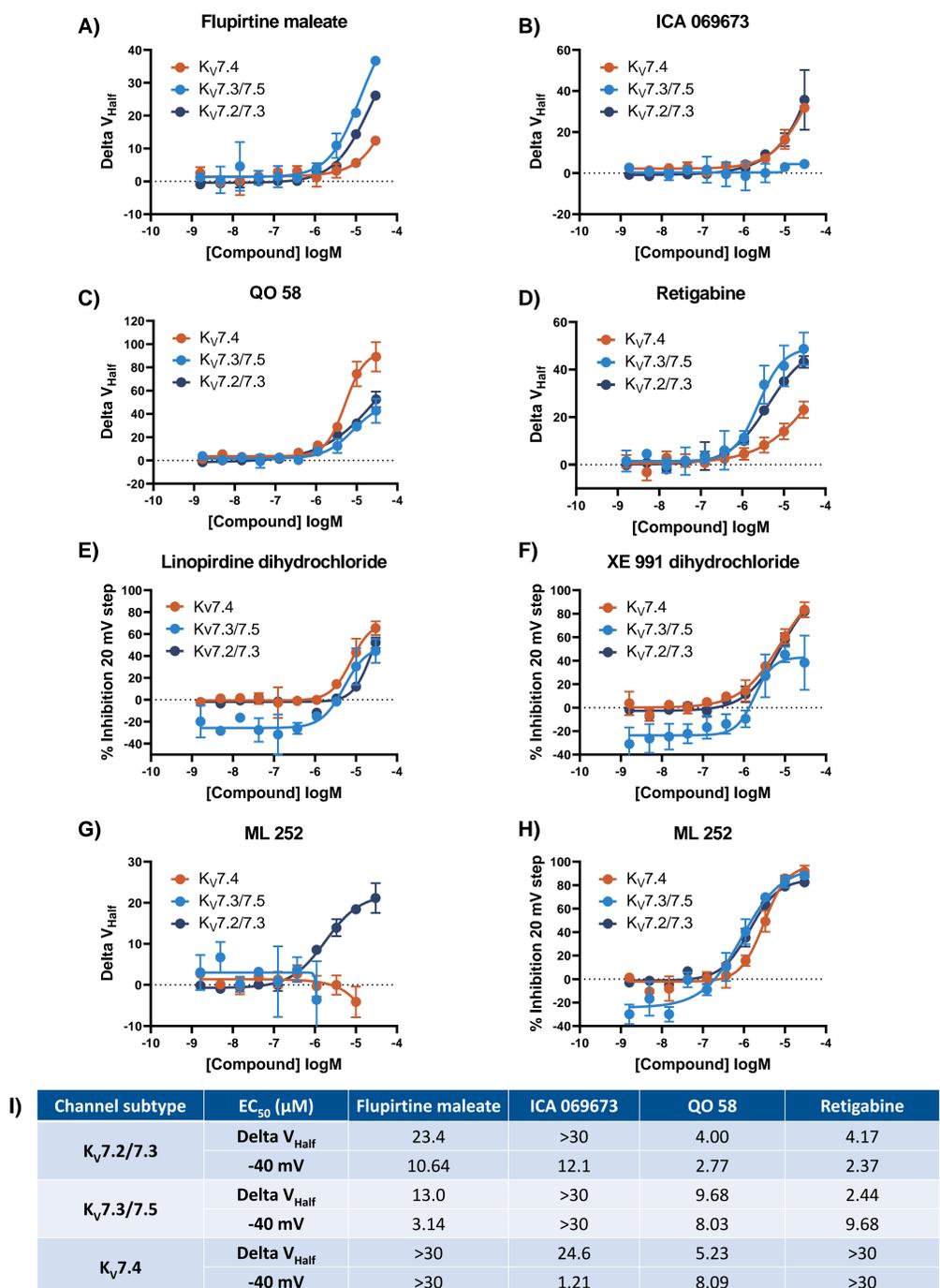


Figure 2. K_v7 pharmacology data.

Example graphs showing reference compound profiling in K_v7 assays, with concentration-response curves from delta V_{Half} and % activation/inhibition at two step voltages (+20 and -40 mV). Reference activators flupirtine maleate (A), ICA 069673 (B), QO 58 (C) and retigabine (D), alongside reference inhibitors Linopirdine (E), XE 991 dihydrochloride (F) and ML 252 (G and H). Summary table showing EC₅₀/IC₅₀ for all compounds tested (I and J).

References

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